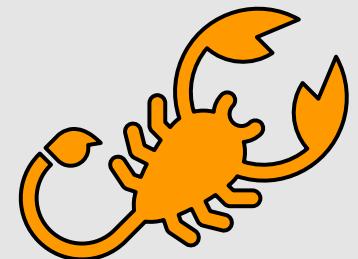


# A Survival Guide to Data Analysis

BERDC Special Topics Talk 15



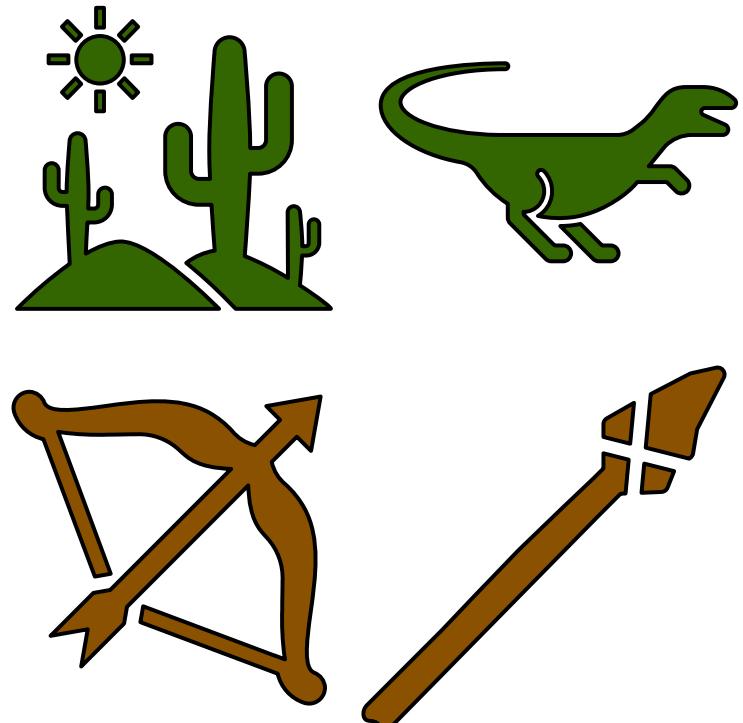
**DaCCoTA**  
DAKOTA COMMUNITY COLLABORATIVE  
ON TRANSLATIONAL ACTIVITY

Dr. Mark Williamson  
Biostatistics, Epidemiology,  
and Research Design Core

# Opening

Goal: Deal with Data Analysis when backed against a wall

- ⚒ Set the stage for dealing with data
- ⚒ State three classes of problems
- ⚒ Discuss three solutions for each problem
- ⚒ Provide examples for each solution

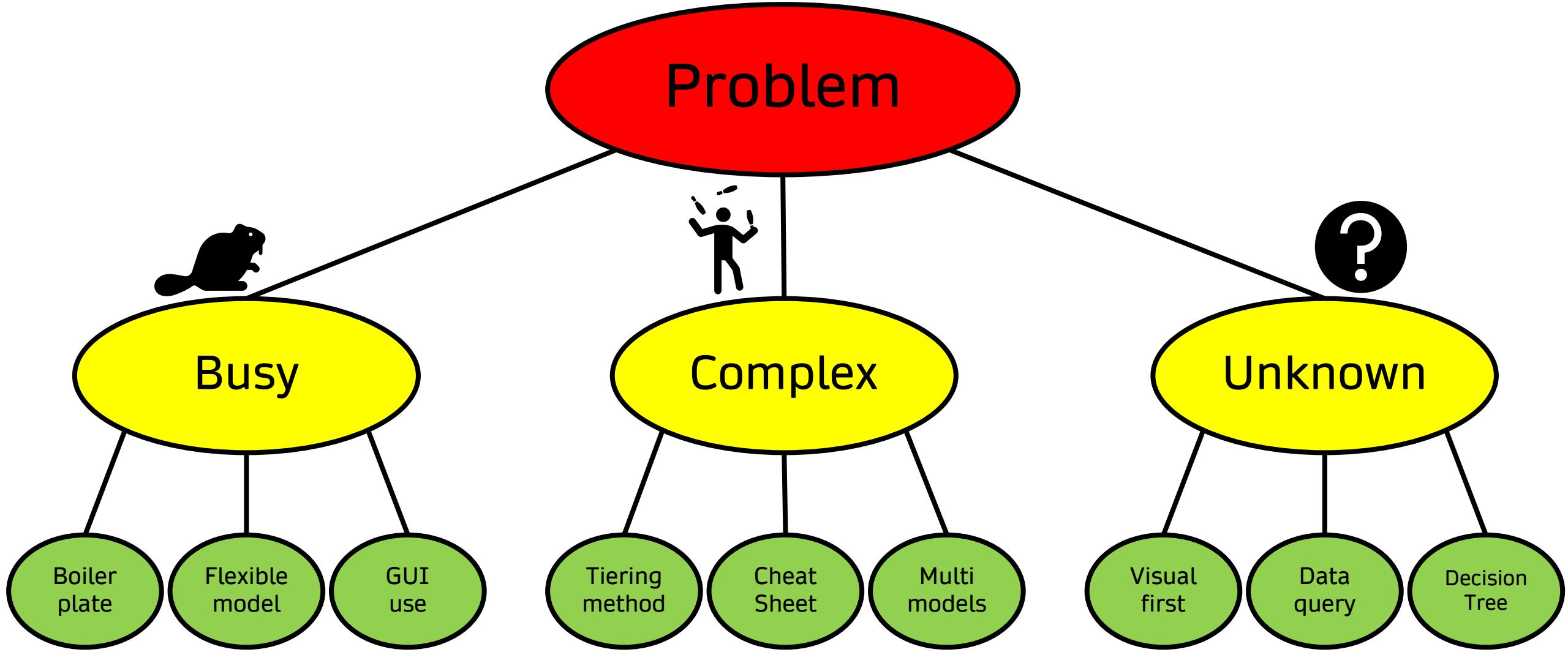


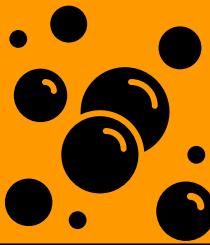
# Introduction

- ❖ Best case scenario for statistical analysis needs is to consult/collaborate with a statistician or statistically-minded researcher
- ❖ Not always possible (funding, location, educational situation, etc.)
- ❖ Take these tools with a grain of salt
- ❖ Steps in this guide are not mutually exclusive



# Content





# Boilerplate

- ❖ Create a standardized document of code that is ready for common procedures and only needs to be filled in with specifics

SAS Coding Tips | 2017

## Sample Comparisons

### T-tests:

```
proc ttest data=one;
  var Var2; *Numeric variable;
  class Var1; *Categorical groups for comparison;
```

### Paired t-test:

```
proc ttest data=one;
  paired Var1 Var2;
```

### Multiple Testing:

```
proc glimmix data=one;
  class Var1; * Categorical groups for comparison;
  model Response = Var1;
  lsmeans Var1 / adjust=TUKEY;
  lsmeans Var1 / adjust=BON;
```

### Non-parametric:

```
proc npar1way data=one;
  var Var2;
  class Var1;
```

### FREQ tables and Chi-square tests:

```
proc freq data = one; BY Experiment;
  weight Count;
  tables Genotype*Disease / chisq;
```

SAS Coding Tips | 2017

## Correlation and Regression

### Correlation between multiple variables, with matrix scatter plot:

```
proc corr data=one plots=matrix;
  var Var1 Var2 Var3 Var4;
```

### Linear Regression:

```
proc reg data=one;
  model depVar = Var2;
```

### Multiple Regression with REG or GLM:

```
proc reg data=one;
  model depVar = Var1;
  model depVar = Var1 Var2; *can include multiple models;
  model depVar = Var1 Var2 Var3;
```

```
proc glm data=one;
  model depVar = Var1|Var2|Var3; *includes all interactions;
```

### Polynomial Regression:

```
data one; set one;
  Var2=Var**2;
proc glm data=one;
  model clutch=length|length2;
```

### Forward and Stepwise selection with GLMSELECT:

```
proc glmselect data=one plots=(Criteria Candidates);
  model response = var1 var2 var3 var4 var5 /
    selection=forward(select=AICC);
proc glmselect data=one plots=(Criteria Candidates);
  model response = var1 var2 var3 var4 var5 /
    selection=stepwise(select=AICC);
```



# Boilerplate

- ❖ Create a standardized document of code that is ready for common procedures and only needs to be filled in with specifics

```
#Simple histogram in ggplot
ggplot(data=DATASET, aes(Y_VAR))+
  geom_histogram(bins=6, fill="blue", col="black")

#Two-sample histogram in ggplot
ggplot(data=DATASET, aes(x=Y_VAR, fill=X_CAT))+ 
  geom_histogram(bins=6, col="black", alpha=0.6, position='identity')+
  scale_fill_manual(values=c("red", "blue"))

#Simple boxplot in ggplot
ggplot(data=DATASET, aes(x=X_CAT, y=Y_VAR)) +
  geom_boxplot()

#Two-way boxplot in ggplot
ggplot(data=DATASET, aes(x=X_CAT1, y=Y_VAR, fill=X_CAT2)) +
  geom_boxplot()
```

```
#Simple scatter plot in ggplot
ggplot(data=DATASET, aes(x=X_NUM, y=Y_VAR)) +
  geom_point()

#Two-way scatter plot in ggplot
ggplot(data=DATASET, aes(x=X_NUM, y=Y_VAR, fill=X_CAT, color=X_CAT)) +
  geom_point()

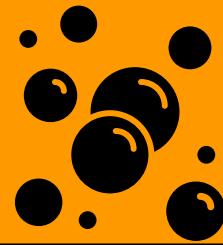
#Spaghetti plot in ggplot
ggplot(data=DATASET, aes(x=X_CAT, y=Y_VAR, group=ID)) +
  geom_line()

#Logistic regression plot in ggplot
ggplot(data=DATASET, aes(x=X_NUM, y=Y_BIN)) +
  geom_point() +
  stat_smooth(method="glm", se=FALSE, method.args=list(family=binomial))

#Bubble plot in ggplot
ggplot(data=DATASET, aes(x=X_NUM1, y=Y_VAR, size=X_NUM2))+ 
  geom_point()
```

```
#Simple bar plot in ggplot
DATASET_sum<-DATASET %>%
  group_by(X_CAT) %>%
  summarise(mean=mean(Y_VAR),
           sd = sd(Y_VAR),
           error = qt(0.975,df=n()-1)*sd/sqrt(n()),
           ul = mean + error,
           ll = mean - error)
ggplot(data=DATASET_sum, aes(x=X_CAT, y=mean, fill=X_CAT)) +
  geom_bar(stat="identity") +
  geom_errorbar(aes(ymin=ll, ymax=ul), width=0.1)

#Two-way bar plot in ggplot
DATASET_sum<-DATASET %>%
  group_by(X_CAT1, X_CAT2) %>%
  summarise(mean=mean(Y_VAR),
           sd = sd(Y_VAR),
           error = qt(0.975,df=n()-1)*sd/sqrt(n()),
           ul = mean + error,
           ll = mean - error)
ggplot(data=DATASET_sum, aes(x=X_CAT1, y=mean)) +
  geom_bar(aes(fill=X_CAT2), stat="identity",
           position=position_dodge()) +
  geom_errorbar(aes(ymin=ll, ymax=ul, group=tension), width=0.1,
           position=position_dodge(0.9))
```



# Boilerplate

- ❖ Create a standardized document of code that is ready for common procedures and only needs to be filled in with specifics

```
barchart.anova <- function(df, cat_var, num_var) {  
  cat_var <- enquo(cat_var)  
  num_var <- enquo(num_var)  
  df_sum<-df %>%  
    group_by (!!cat_var) %>%  
    summarise(mean=mean (!!num_var),  
             sd = sd (!!num_var),  
             error = qt(0.975,df=n()-1)*sd/sqrt(n()),  
             ul = mean + error,  
             ll = mean - error)  
  #print(df_sum)  
  df_plot <- ggplot(df_sum, aes (!!cat_var, mean, fill= !!cat_var))+  
    geom_bar(stat="identity", color="black") +  
    geom_hline(yintercept = 0) +  
    geom_errorbar(aes(ymin=ll, ymax=ul), width=0.1) +  
    theme_classic() + theme(legend.position="none")  
}
```

```
#Example  
barchart.anova(iris, Species, Sepal.Length)  
#basic  
df_plot  
#customized  
df_plot + labs(y="Sepal length (cm", x="Species") +  
  scale_y_continuous(limits=c(0,7),breaks=c(0:7))+  
  geom_text(aes(label=c('*', '*', '*')), vjust=-0.8, size=6)
```



# Flexible Models

- ❖ Use flexible models that can run multiple tests just by tweaking parameters, rather than having to run different code for each and every test



## #FLEXIBLE MODELS

```
head(mtcars)
```

```
t.model <- lm(mpg~as.factor(vs), data=mtcars)  
summary(t.model)
```

```
a.model <- lm(mpg~as.factor(carb), data=mtcars)  
summary(a.model)
```

```
r.model <- lm(mpg~wt, data=mtcars)  
summary(r.model)
```



# Flexible Models

- ❖ Use flexible models that can run multiple tests just by tweaking parameters, rather than having to run different code for each and every test

```
PROC GLIMMIX data=sashelp.Class;  
  class Sex;  
  model Height=Sex /solution dist=normal;  
  lsmeans Sex /cl;  
  ods output LSmeans=Class_lsm;  
  
PROC SGPlot data=Class_lsm;  
  vbarparm category=Sex response=Estimate/limitupper=Upper  
  limitlower=Lower;
```

T-test

```
PROC GLIMMIX data=sashelp.bweight;  
  class MomEdLevel;  
  model Weight=MomEdLevel/solution dist=normal;  
  lsmeans MomEdLevel / cl;  
  ods output LSMeans=Bweight_lsm;  
  
PROC SGPlot data=Bweight_lsm;  
  vbarparm category=MomEdLevel  
  response=Estimate/  
  limitupper=Upper limitlower=Lower;
```

ANOVA

```
PROC GLIMMIX data=sashelp.bmimen;  
  model BMI=age/solution dist=normal;  
  output out=Bmimen_pred pred lcl ucl;  
  
PROC SGPlot data=Bmimen_pred;  
  band x=age lower=lcl upper=ucl;  
  scatter x=age y=BMI;  
  series x=age y=Pred;
```

Regression

```
PROC GLIMMIX data=multicenter;  
  class center group;  
  model SideEffect/n = group / solution;  
  random center;  
  lsmeans group / ilink cl;  
  ods output LSMeans=lsm1;  
  
PROC SGPlot data=lsm1;  
  vbarparm category=group  
  response=Mu /  
  limitlower=LowerMu  
  limitupper=UpperMu;
```

Generalized  
Linear Mixed  
Model



# GUI Use

## ❖ Use a Graphical Use Interface to rapidly select and run tests

The image displays three screenshots of the R Commander graphical user interface:

- Screenshot 1 (Top Left):** Shows the "Select Data Set" dialog box. The "chickwts" dataset is selected in the list, with "OK" highlighted.
- Screenshot 2 (Bottom Left):** Shows the main R Commander window with the "Data" menu open. The "Manage variables in active data set..." option is highlighted, showing a submenu with various variable manipulation options like "Recode variables...", "Compute new variable...", and "Delete variables from data set ...".
- Screenshot 3 (Right):** Shows the main R Commander window with the "Graphs" menu open. The "Plot of means..." option is highlighted, showing a submenu with other plot types such as "Index plot...", "Dot plot...", "Histogram...", and "Scatterplot...".

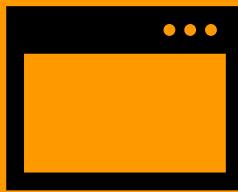


# GUI Use

❖ Use a Graphical Use Interface to rapidly select and run tests

The image shows a screenshot of the DaCCoTA software interface. On the left, there are two overlapping dialog boxes for 'Plot Means'. The top dialog is the 'Data' tab, showing 'Factors (pick one or two)' set to 'feed' and 'Response Variable (pick one)' set to 'weight'. The bottom dialog is the 'Options' tab, showing 'Error Bars' set to 'Confidence intervals' (Level of confidence: 0.95), 'Plot Labels' with 'x-axis label' as 'Feed Type' and 'y-axis label' as 'Chick Weight', and 'Position of Legend' set to 'To right of graph'. Both dialogs have 'OK', 'Cancel', and 'Apply' buttons. To the right of these dialogs is a plot titled 'Plot of Means' showing Chick Weight (Y-axis, 150 to 350) versus Feed Type (X-axis, casein, horsebean, linseed, meatmeal, soybean, sunflower). The plot displays mean weights with 95% confidence interval error bars. The data points are approximately: casein (~325), horsebean (~160), linseed (~220), meatmeal (~280), soybean (~250), and sunflower (~330).

Feed Type	Chick Weight (approx.)
casein	325
horsebean	160
linseed	220
meatmeal	280
soybean	250
sunflower	330



# GUI Use

❖ Use a Graphical Use Interface to rapidly select and run tests

The screenshot shows the R Commander interface. The 'Statistics' menu is open, and the 'Means' option is selected. Within the 'Means' submenu, 'One-way ANOVA...' is highlighted with a blue selection bar. The R console area shows some code related to the chickwts dataset.

```
Rcmdr> summary(AnovaModel.1)
Df Sum Sq Mean Sq F value Pr(>F)
feed      5 231129  46226  15.37 5.94e-10 ***
Residuals 65 195556   3009
---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Rcmdr> with(chickwts, numSummary(weight, groups=feed, statistics=c("mean", "sd")))
    mean     sd data:n
casein 323.5833 64.43384   12
horsebean 160.2000 38.62584   10
linseed 218.7500 52.23570   12
meatmeal 276.9091 64.90062   11
soybean 246.4286 54.12907   14
sunflower 328.9167 48.83638   12
```



# GUI Use

❖ Use a Graphical Use Interface to rapidly select and run tests

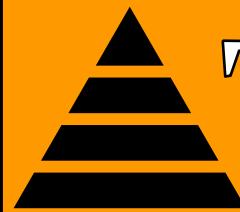
The screenshot shows the IBM SPSS Statistics Data Editor interface. The main window displays a data view of 20 rows and 12 columns. The columns include age, ed, employ, address, income, debtinc, creddebt, othdebt, default (dependent variable), preddef1, preddef2, and predc. The 'default' column contains values like 'Yes' and 'No'. The 'ed' column shows levels of education such as 'Some college', 'Did not complete high school', and 'High school degree'. The 'income' column shows values ranging from 17 to 176.00. The 'debtinc' column shows values like 9.30 and 11.36. The 'preddef1' and 'preddef2' columns show numerical values. The 'predc' column shows values like .80839, .78864, etc.

A 'Logistic Regression' dialog box is open in the foreground. It has the following settings:

- Dependent:** Previously defaulted [default]
- Covariates:** age, ed, employ, address, income
- Method:** Enter

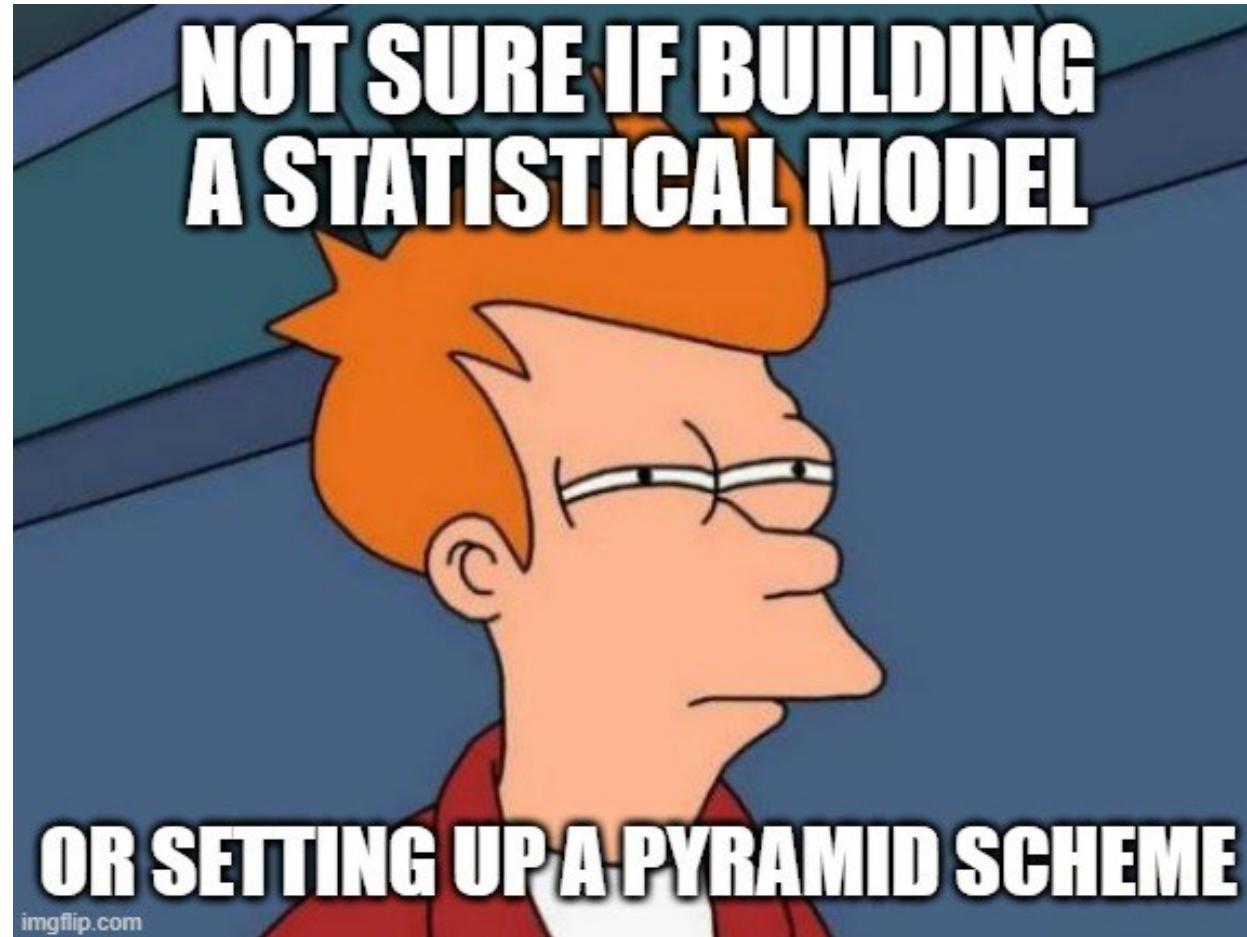
Buttons in the dialog box include OK, Paste, Reset, Cancel, and Help.

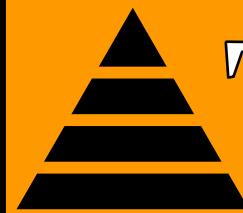
The bottom of the screen shows the status bar with 'IBM SPSS Statistics Processor is ready' and 'Unicode:ON'.



# Tiering Method

- ⚒ Start with simplest model design and progressively build from there

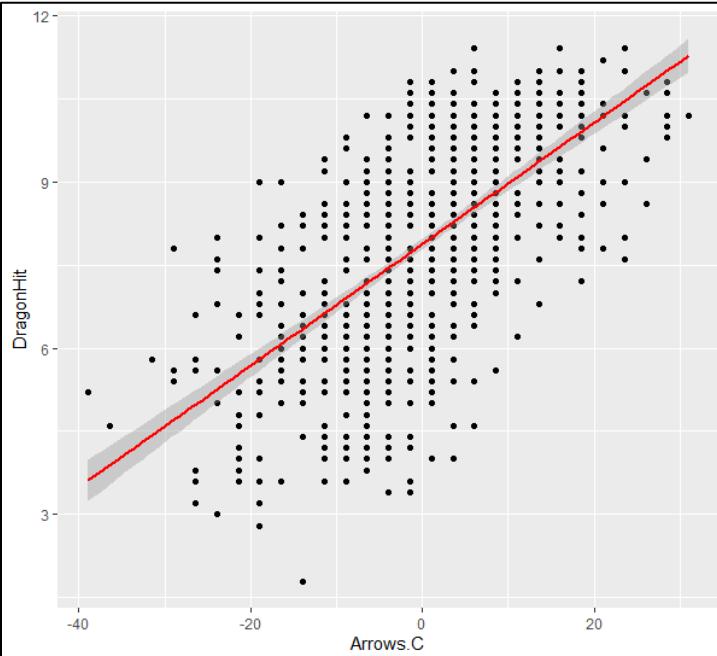




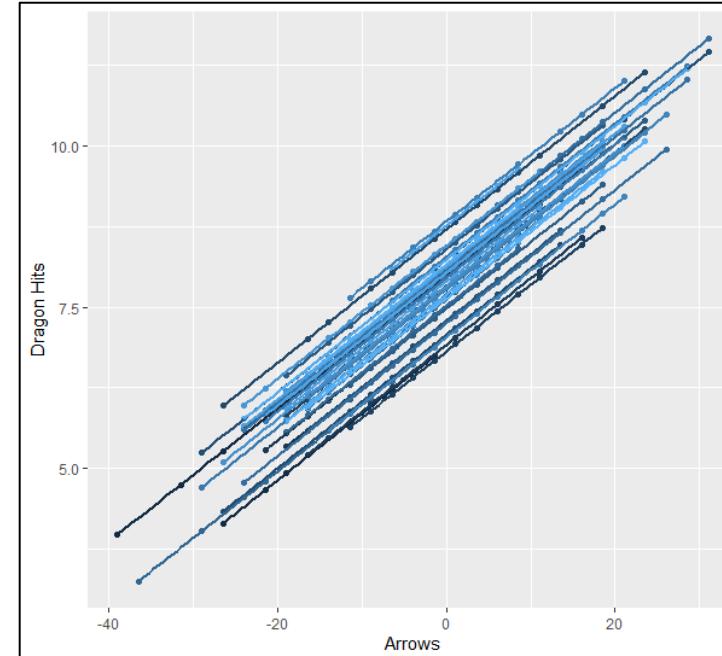
# Tiering Method

- ⚒ Start with simplest model design and progressively build from there

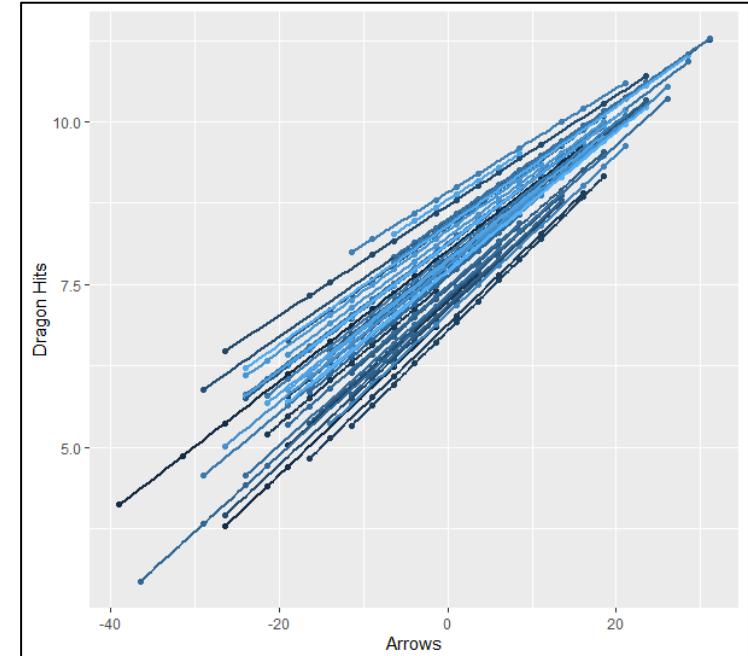
`D3_M1 <- lm(DragonHit ~ Arrows.C,  
data=Dragon2)`



`D3_M2 <- lmer(DragonHit ~ Arrows.C  
+ (1 | Cathedral),  
data=Dragon2, REML=F)`



`D3_M3 <- lmer(DragonHit ~ Arrows.C  
+ (Arrows.C | Cathedral),  
data=Dragon2, REML=F)`



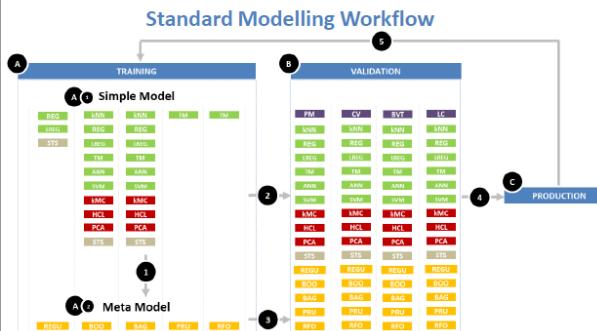


# Cheat Sheet

❖ Deploy sheets with quickly available information for statistical methods, especially more advanced concepts

## Machine Learning Modelling in R : : CHEAT SHEET

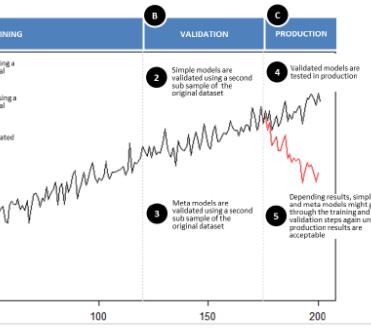
Supervised & Unsupervised Learning		
ALGORITHM	DESCRIPTION	R PACKAGE::FUNCTION
Naive Bayes classifier	A classification technique based on Bayes' theorem with an assumption of independence among predictors. In simple terms, a class is associated with a particular feature in a dataset. The probability of presence or absence of a particular feature in a class is calculated.	naiveBayes
k-Nearest Neighbors	A non parametric method used for classification and regression. In both cases, the input consists of the k closest training examples in the feature space. The output is based on majority vote of its neighbors (classification) or weighted average of the neighbors (regression).	knn
Linear Regression	Model the linear relationship between a scalar dependent variable Y and one or more explanatory variables (or independent variables), usually X.	lm(dist ~ speed, data = cars)
Logistic Regression	The idea is to consistently divide (strand) the training dataset based on the input features until an assignment rule (e.g. "if the first variable is 0, then 0") is reached.	glm(Y ~ ., family = binomial(link = "logit"), data = X)
Tree-Based Models	Tree-based models are built from units called "decision trees". A decision tree is a flowchart-like structure in which each node represents an input variable, each branch represents a decision rule, and each leaf node represents the outcome.	rpart(rpart ~ Age + Number ~ Start, data = lymphoma)
Artificial Neural Network	Neural networks are built from units called "perceptrons". A perceptron is a unit that takes multiple inputs, applies a weight vector to them, adds a bias, and then applies an activation function to produce an output.	neuralnet(neuralnet ~ I(data\$train[, -id]), linear.out = TRUE)
Support Vector Machine	A data classification method that separates data using hyperplanes.	svm(svm ~ I(data\$train[, -id]), kernel = "linear", type = "C-classification", cost = 1000)
Principal Component Analysis	A procedure that uses an orthogonal transformation to convert a set of observations of possibly correlated variables into a set of values of linearly uncorrelated variables called principal components.	prcomp(prcomp ~ I(data\$train[, -id]), center = TRUE, scale = TRUE)
k-Means Clustering	Algorithms at partitioning observations into k clusters in which each observation belongs to the cluster with the nearest mean.	kmeans(kmeans ~ I(data\$train[, -id]), centers = 3, iter.max = 30, start = 1, algorithm = "Hartigan-Wong", "Lloyd", "MacQueen")
Hierarchical Clustering	An approach that builds a hierarchy from the bottom up, where the number of clusters to be specified beforehand.	hc(hc ~ I(data\$train[, -id]), members = N(3))



## Meta-Algorithm, Time Series & Model Validation

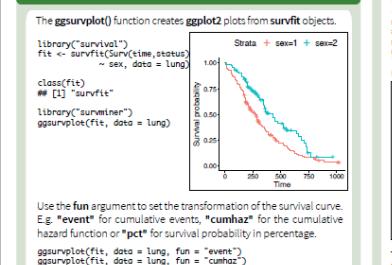
ALGORITHM	DESCRIPTION	R PACKAGE::FUNCTION
Regularization (L1/L2/ridge)	Regularization adds a penalty on the different parameters of a model to prevent overfitting. L1 regularization is useful for feature selection, while L2 regularization is useful for generalization.	glmnet(glmnet ~ I(data\$train[, -id]), family = "gaussian", alpha = 1, lambda = 0.001)
Boosting	A process of iteratively refitting, e.g. by reweighting, of estimated regression or classification functions through their residuals so as to obtain better fits.	gbm(gbm ~ I(data\$train[, -id]), family = "gaussian", n.trees = 100)
Bagging	Bagging is a way to increase the power of a single classifier by taking multiple random samples (with replacement) of the training data set, and each of which is used to train a classifier. The final prediction is the original test set.	foreach(d = data.frame(1:10, y = norm(10)), gbm(~ ., data = d))
Pruning	Pruning is a technique that reduces the complexity of a decision tree by removing parts of the tree that have little or no power of classification. Pruning reduces the complexity of the final model, thus improving the accuracy by reducing overfitting.	prune(prune ~ I(data\$train[, -id]), cp = 0.1)
Random Forest	An ensemble learning method that constructs multiple decision trees for classification and regression. The final output is determined by averaging the outputs of the individual trees.	randomForest(randomForest ~ I(data\$train[, -id]), y = subset, na.action = na.omit)
Random Sampling	Random sampling of observations for training testing a model. This is useful for dealing with large datasets. Random sampling may either destroy serial correlation or introduce it in data which it would not be explicit.	strata(strata ~ I(data\$train[, -id]), sample = TRUE)
Performance Metrics	Depends on the problem: • Regression: squared errors, outliers, error rate, etc. • Classification: accuracy, precision, recall, f-score, etc.	autoCorrelation(autoCorrelation ~ I(data\$train[, -id]), type = "lagged", lags = 10) statistic(statistic ~ I(data\$train[, -id]), type = "KS") confusionMatrix(confusionMatrix ~ I(data\$train[, -id]), type = "table")
Statistical Variance Tradeoff	• Simple models with few parameters are easier to compute but may lead to poorer fits (high bias). • Complex models with many parameters can estimate fits but may over fit (high variance).	ggplot2::autoplot(gbm, center = TRUE, scale = FALSE, n = 100, plot_type = "OOB")
Cross-validation	Cross-validation compares the performance of different models with different numbers of values or types of parameters.	caret::createDataPartition(caret::createDataPartition ~ I(data\$train[, -id]), method = "stratified", list = TRUE, p = 0.8, times = 100)
Learning Curves	Learning curves plot a model's training and test errors, or the model's performance metric, depending on the training set size.	caret::learning_curve ~ I(data\$train[, -id])

## Time Series View



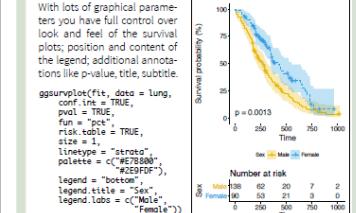
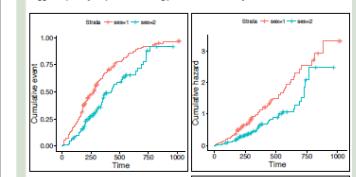
## Creating Survival Plots Informative and Elegant with survminer

### Survival Curves



Use the `fun` argument to set the transformation of the survival curve. E.g. `"event"` for cumulative events, `"cumhaz"` for the cumulative hazard function or `"pctsurv"` for survival probability in percentage.

`ggsurvplot(fit, data = lung, fun = "event")`  
`ggsurvplot(fit, data = lung, fun = "cumhaz")`



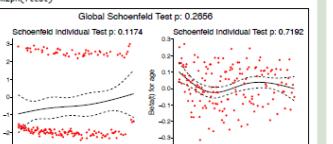
This one pager presents the `survminer` package [Alboukadel Kassambara, Marcin Kosinski, Przemyslaw Biecek, Scheipl Fabian 2021] in version 0.4.9. See <https://github.com/kassambara/survminer/> for more details.

### Diagnostics of Cox Model

The function `cox.zph()` from `survival` package may be used to test the proportional hazards assumption for a Cox regression model fit. The graphical verification of this assumption may be performed with the function `ggsurvph()` from the `survminer` package. For each covariate it produces plots with scaled Schoenfeld residuals against the time.

`library(survival)`  
`lungSph <- ifelse(lung$age > 70, ">70", "<=70")`  
`fit <- coxph(Surv(time, status) ~ sex + ph.ecog + age, data = lung)`  
`fit`

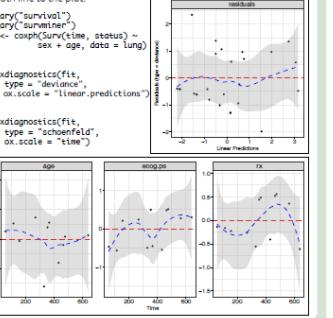
`# Call:`  
`#> coxph(formula = Surv(time, status) ~ sex + ph.ecog + age, data = lung)`  
`#> #> sex: sex chisq 0.117`  
`#> sex: age exp(coef)* se(coef)^2 5 0.00002`  
`#> ph.ecog: sex0.875 0.129 0.719`  
`#> ph.ecog: age0.651 0.266`  
`library("survminer")`  
`ggsurvph(fit)`



The function `gcoxdiagnostics()` plots different types of residuals as a function of time, linear predictor or observation id. The type of residual is selected with `type` argument. Possible values are "martingale", "deviance", "score", "schoenfeld", "dfbeta", "dfbetas", and "scaleds". The `ox.scale` argument defines what shall be plotted on the OX axis. Possible values are "linearPredictions", "observation.id", "time". Logical arguments `hline` and `cline` may be used add horizontal line or smooth line to the plot.

`library("survival")`  
`library("survminer")`  
`lungSph <- ifelse(lung$sex == 1, "Male", "Female")`  
`fit <- coxph(Surv(time, status) ~ sex + age, data = lung)`

`gcoxdiagnostics(fit, type = "deviance", ox.scale = "linear.predictions")`  
`gcoxdiagnostics(fit, type = "schoenfeld", ox.scale = "time")`



Note that it is not necessary to include the grouping factor in the Cox model. Survival curves are estimated from Cox model for each group defined by the factor independently.

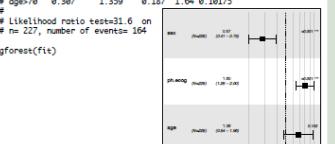
`lungAge3 <- cut(lung$age, c(35, 55, 65, 85))`  
`gcoxdiagnostics(fit, data = lung, variable = "age3")`

### Summary of Cox Model

The function `gforest()` from the `survminer` package creates a forest plot for a Cox regression model fit. Hazard ratio estimates along with confidence intervals and p-values are plotted for each variable.

`library("survival")`  
`library("survminer")`  
`lungSph <- ifelse(lung$age > 70, ">70", "<=70")`  
`fit <- coxph(Surv(time, status) ~ sex + ph.ecog + age, data = lung)`  
`fit`

`# Call:`  
`#> coxph(formula = Surv(time, status) ~ sex + ph.ecog + age, data = lung)`  
`#> #> sex: sex chisq 0.117`  
`#> sex: age exp(coef)* se(coef)^2 5 0.00002`  
`#> ph.ecog: sex0.875 0.129 0.719`  
`#> ph.ecog: age0.651 0.266`  
`library("survminer")`  
`gforest(fit)`

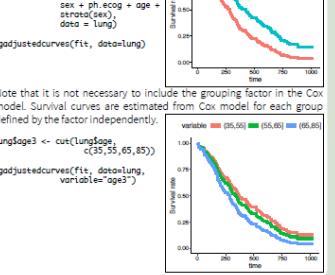


The function `gadjustedcurves()` from the `survminer` package plots Adjusted Survival Curves for Cox Proportional Hazards Model. Adjusted Survival Curves show how a selected factor influences survival estimated from a Cox model.

Note that these curves differ from Kaplan Meier estimates since they present expected survival based on given Cox model.

`library("survival")`  
`library("survminer")`  
`lungSex <- ifelse(lung$sex == 1, "Male", "Female")`  
`fit <- coxph(Surv(time, status) ~ sex + age, data = lung)`

`gadjustedcurves(fit, data = lung, variable = "sex")`



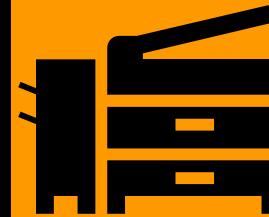
CC BY SA Arnaud Amsellem • [thertrader@gmail.com](mailto:thertrader@gmail.com) • [www.thertrader.com](http://www.thertrader.com) • Updated: 2018-03



# Cheat Sheet

❖ Deploy sheets with quickly available information for statistical methods, especially more advanced concepts

Examples	STATA	SPSS	Excel	SAS	R
Summary statistics	Data->Describe Data-> Summary Statistics OR summarize <i>num_var</i>	Analyze -> Descriptive Statistics -> Descriptives	=AVERAGE( <i>num_var</i> ) =MEDIAN( <i>num_var</i> ) =STDEV.S( <i>num_var</i> ) ...	PROC UNIVARIATE; var <i>num_var</i> ;	summary( <i>num_var</i> )
Histogram	Graphics-> Histogram OR histogram <i>num_var</i>	Graphs -> Chart Builder -> Histogram	Insert (Charts)-> Histogram	PROC SGPLOT; histogram <i>num_var</i> ;	hist( <i>num_var</i> )
Boxplot	Graphics-> Box plot OR graph box <i>num_var</i> , over( <i>cat_var</i> )	Graphs -> Chart Builder -> Boxplot	Insert (Charts)-> Box and Whisker	PROC SGPLOT; vbox <i>num_var</i> / group= <i>cat_var</i> ;	plot( <i>num_var</i> ~ <i>cat_var</i> )
Bar plot	Graphics-> Bar Chart OR graph bar (mean) <i>num_var</i> , over( <i>cat_var</i> )	Graphs -> Chart Builder -> Bar	Insert (Charts)-> Column	PROC SGPLOT; vbarparm category= <i>cat_var</i> treatment= <i>num_mean</i> ;	means <- c( <i>mean_cat1</i> , <i>mean_cat2</i> ) barplot(means)
Scatterplot	Graphics -> Two-way graph OR twoway (scatter <i>num_var1</i> <i>num_var2</i> )	Graphs -> Chart Builder -> Scatter/Dot	Insert (Charts)-> Scatter	PROC SGPLOT; Scatter y= <i>num_var1</i> x= <i>num_var2</i> ;	plot( <i>num_var1</i> , <i>num_var2</i> )
T-test	Statistics -> Summaries, tables, and tests -> Classical tests of hypotheses ->t tests OR ttest <i>num_var</i> , by( <i>cat_var</i> )	Analyze -> Compare means-> Independent-Samples T Test	=TTEST( <i>num_var1</i> , <i>num_var2</i> , tails, type)	PROC TTEST; var <i>num_var</i> ; class <i>cat_var</i> ;	t.test( <i>num_var</i> ~ <i>cat_var</i> )
ANOVA	Statistics-> Linear models and related -> ANOVA/MANOVA -> One-way ANOVA OR oneway <i>num_var</i> <i>cat_var</i>	Analyze -> Compare means-> One-Way ANOVA	Data Analysis (add-on) -> Anova: Single Factor	PROC ANOVA; class <i>cat_var</i> ; model <i>num_var</i> = <i>cat_var</i> ;	aov( <i>num_var</i> ~ <i>cat_var</i> )
Normal linear regression model	Statistics-> Linear models and related -> Linear regression OR regress <i>num_var1</i> <i>num_var2</i>	Analyze -> Regression-> Linear	Data Analysis (add-on) -> Regression	PROC REG; model <i>num_var1</i> = <i>num_var2</i> ;	lm( <i>num_var1</i> ~ <i>num_var2</i> )
Logistic regression model	Statistics-> Binary outcomes-> Logistic regression OR logit <i>binary_var</i> <i>num_var</i>	Analyze -> Regression-> Binary Logistic	N/A	PROC LOGISTIC; model <i>event/trial</i> = <i>num_var2</i> ;	glm( <i>binary_var</i> ~ <i>num_var</i> , family=binomial)
Poisson regression model	Statistics -> Count outcomes-> Poisson regression OR Poisson <i>count_var</i> <i>num_var</i>	Analyze -> Regression-> Generalized Linear Models	N/A	PROC GLIMMIX; model <i>count_var</i> = <i>num_var</i> /dist=Poisson;	glm( <i>count_var</i> ~ <i>num_var</i> , family=Poisson)
Generalized linear mixed model	Statistics -> Multilevel mixed-effects models -> Generalized linear model OR mglm <i>var1</i> <i>var2</i>    <i>rand_var_eqn</i> , family( <i>distribution</i> ) link( <i>link_function</i> )	Analyze-> Mixed Models-> Generalized Linear	N/A	PROC GLIMMIX; class <i>cat_var</i> ; model <i>num_var1</i> = <i>num_var2</i> <i>cat_var</i> <i>rand_var</i> ; random <i>rand_var</i> ;	Package lme4 lmer( <i>num_var1</i> ~ <i>num_var2</i> + <i>cat_var</i> +(1  <i>rand_var</i> )



# Multi Models

⚒ Be ready to run multiple models that could fit the data and compare best fit

```
btb_lmer1 <- lmer(bdi ~ bdi.pre + time + treatment + drug + length + (1 | subject),  
                   data = BtheB_long, REML=FALSE, na.action = na.omit) #Rand Int
```

```
btb_lmer2 <- lmer(bdi ~ bdi.pre + time + treatment + drug + length + (time | subject),  
                   data = BtheB_long, REML=FALSE, na.action = na.omit) #Rand Int & Slope
```

```
btb_gee1 <- gee(bdi ~ bdi.pre + treatment + length + drug,  
                  data = BtheB_long, id = subject, family = gaussian, corstr = "independence")
```

```
btb_gee2 <- gee(bdi ~ bdi.pre + treatment + length + drug,  
                  data = BtheB_long, id = subject, family = gaussian, corstr = "exchangeable")
```

```
anova(btb_lmer1, btb_lmer2, btb_gee1, btb_gee2)
```

```
PROC GLIMMIX data=DATASET method=RSPL;  
  class RAND1;  
  model Y_VAR=X_VAR1 | X_VAR2;  
  random RAND1;
```



# Multi Models

- ⚒ Be ready to run multiple models that could fit the data and compare best fit

```
reg1 <- lm(Dep ~ Ind1 + Ind2 + Ind3 + Ind4)
sreg1 <- step(reg1, direction="forward")
sreg2 <- step(reg1, direction="backward")
sreg3 <- step(reg1, direction="both")
```

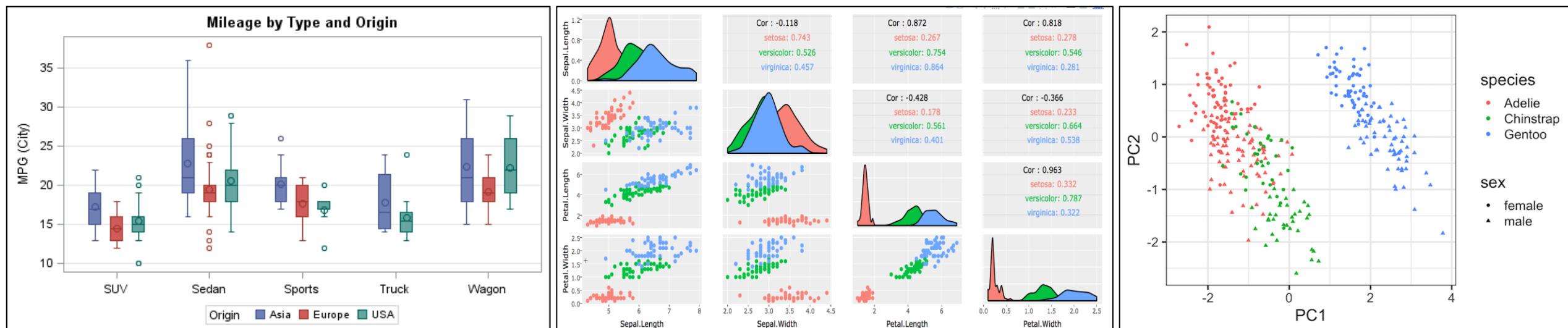
```
PROC GLMSELECT data=sashelp.baseball
plots=all;
class league division;
model logSalary = nAtBat nHits nHome nRuns
nRBI nBB yrMajor crAtBat
crHits crHome crRuns crRbi
crBB league division nOuts
nAssts nError
/ details=all stats=all;
```





# Visual First

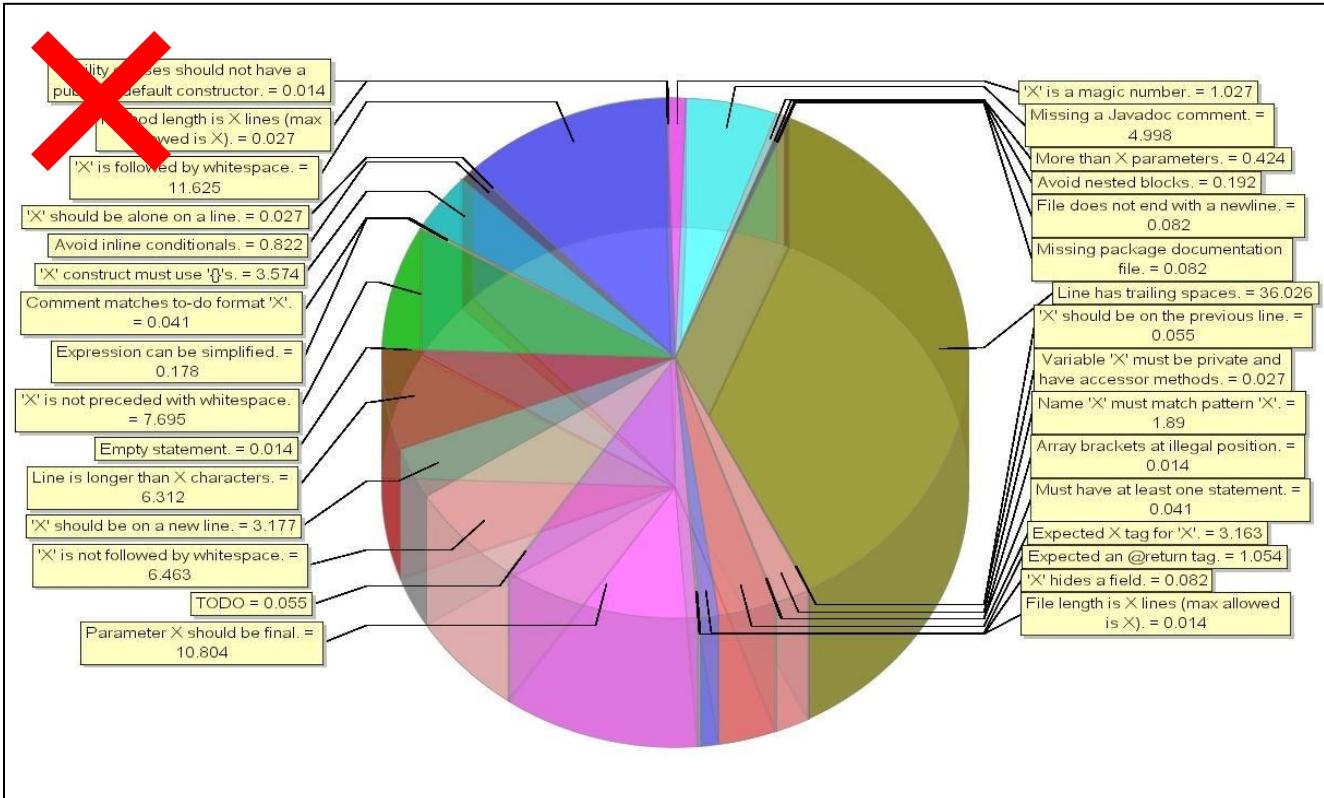
- ⚒ Get to know your data through exploratory data visualization (summary tables, boxplots, scatter plots, dimensional reduction, etc.)





# Visual First

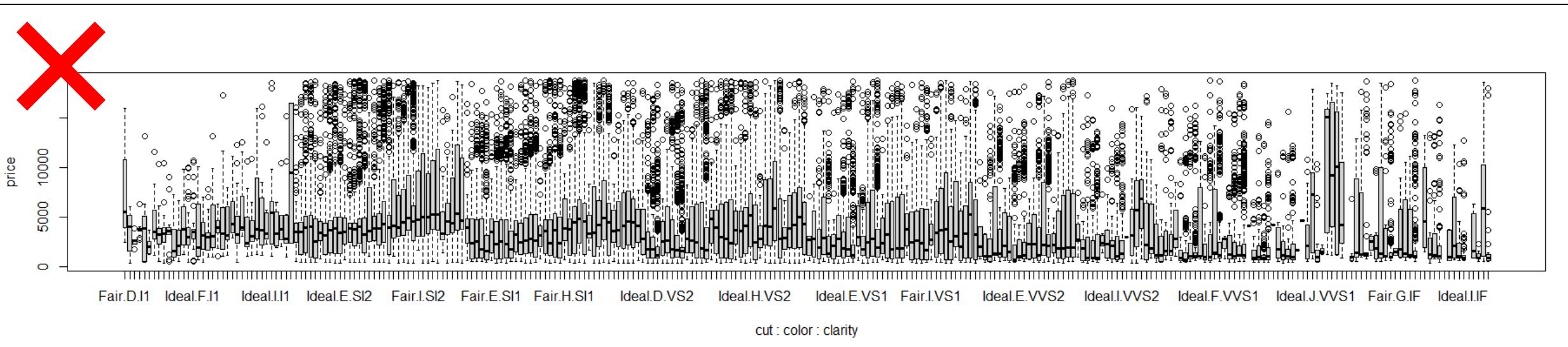
- ❖ Get to know your data through exploratory data visualization (summary tables, boxplots, scatter plots, dimensional reduction, etc.)





# Visual First

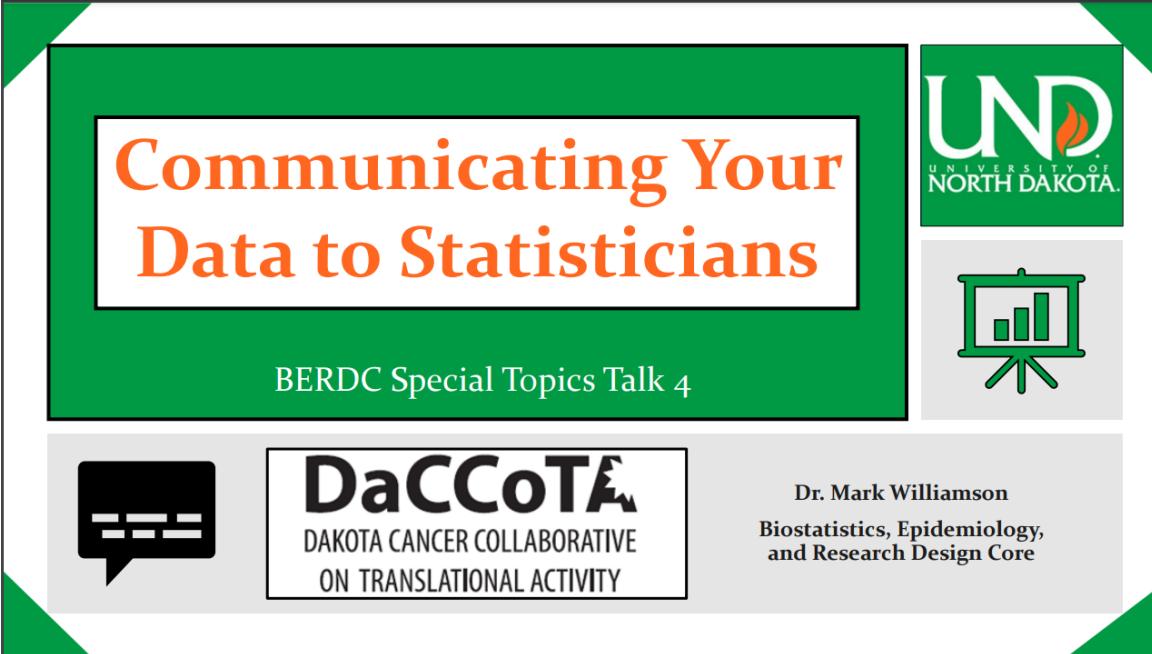
- ⚒ Get to know your data through exploratory data visualization (summary tables, boxplots, scatter plots, dimensional reduction, etc.)





# Data Query

- ❖ Answer questions to get you thinking about your data to figure out structure and variable information



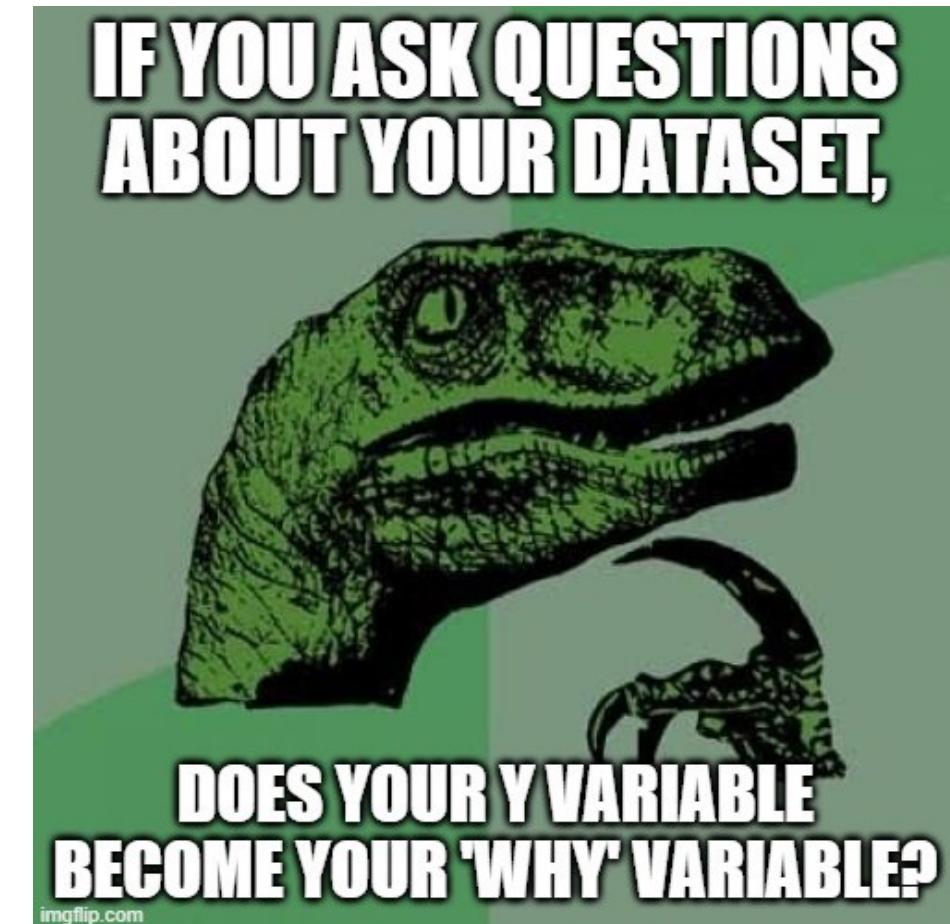
**Communicating Your Data to Statisticians**

BERDC Special Topics Talk 4

 **DaCCoTA**  
DAKOTA CANCER COLLABORATIVE  
ON TRANSLATIONAL ACTIVITY

 Dr. Mark Williamson  
Biostatistics, Epidemiology,  
and Research Design Core

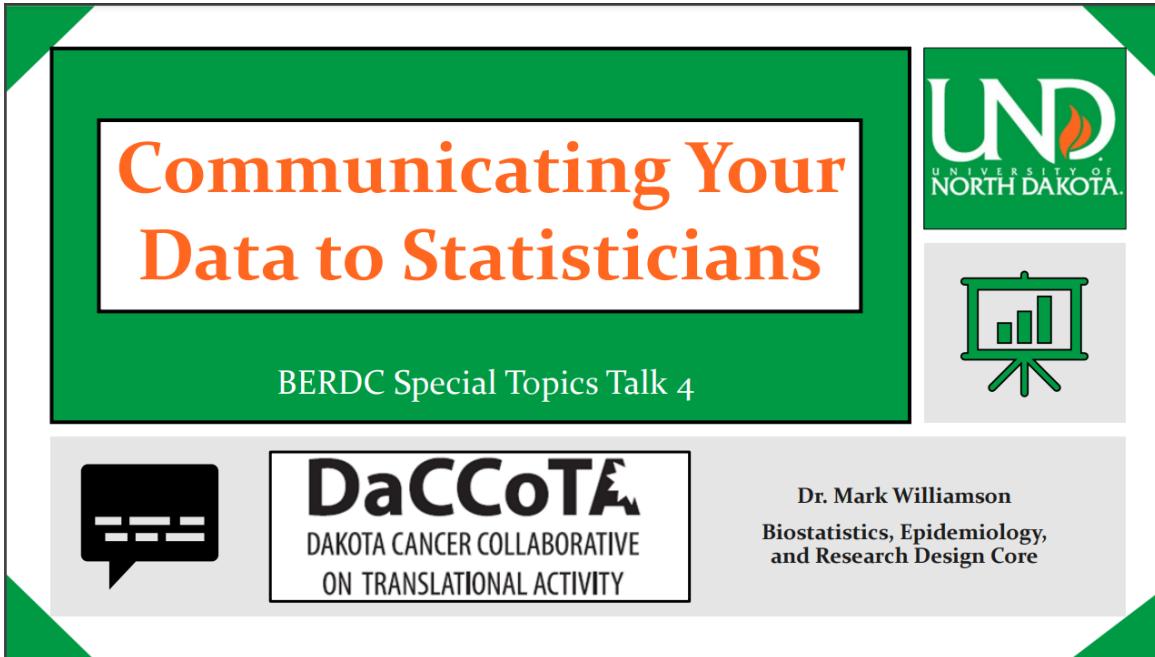
[https://und.qualtrics.com/jfe/form/SV\\_dcyUSPLhD4cmP5Q](https://und.qualtrics.com/jfe/form/SV_dcyUSPLhD4cmP5Q)





# Data Query

- ❖ Answer questions to get you thinking about your data to figure out structure and variable information



**Communicating Your Data to Statisticians**

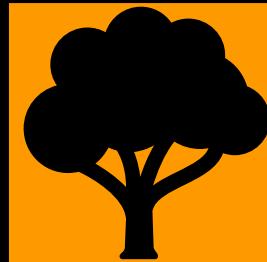
BERDC Special Topics Talk 4

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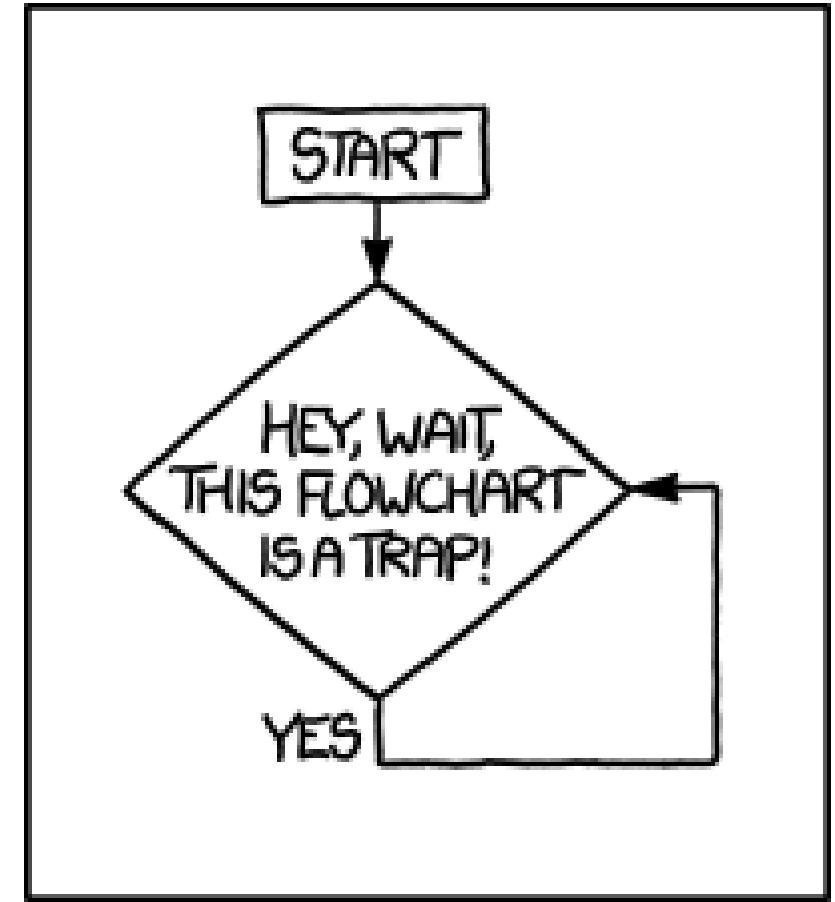
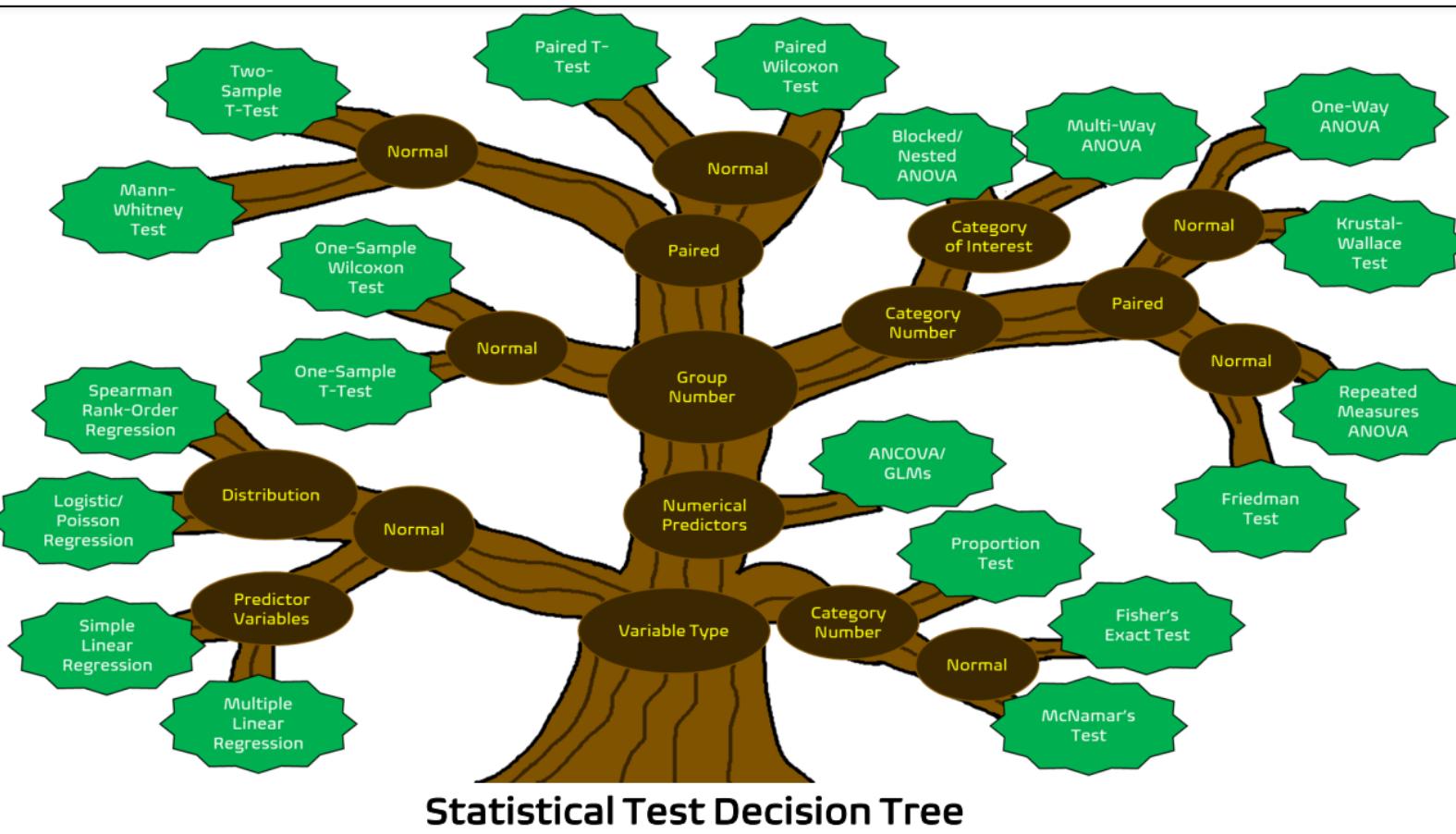
#	Question
1	What is your <b>Y variable</b> ?
2	How is your <b>Y variable measured</b> (categorical-> ordinal or nominal, numerical -> continuous or discrete)?
3	What are your <b>X variables</b> ?
4	How are your <b>X variables measured</b> (categorical-> ordinal or nominal, numerical -> continuous or discrete)?
5	Are their <b>major considerations</b> you need to be aware of (confounding factors, longitudinal design, missing data, etc.)?

[https://und.qualtrics.com/jfe/form/SV\\_dcyUSPLhD4cmP5Q](https://und.qualtrics.com/jfe/form/SV_dcyUSPLhD4cmP5Q)



# Decision Tree

⚒ Walk through branching decisions to determine appropriate test to use



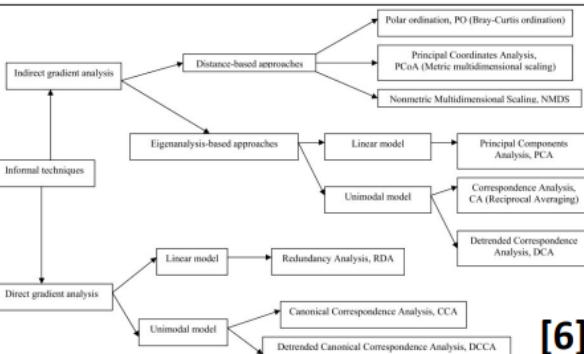
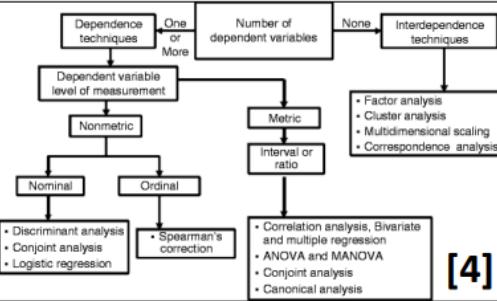


# Decision Tree

⚒ Walk through branching decisions to determine appropriate test to use

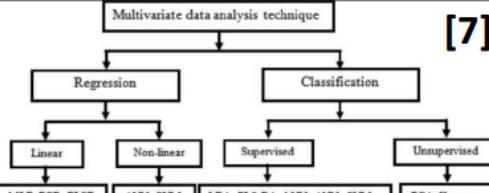
## Classification Problems

[4]

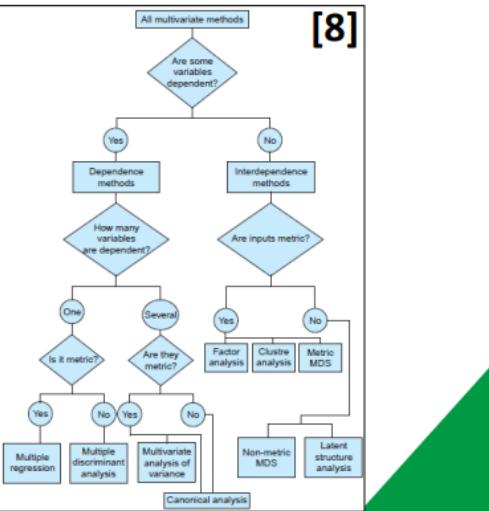


[6]

[7]



[8]



[5]

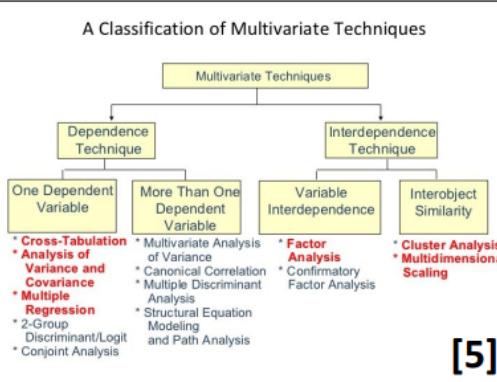
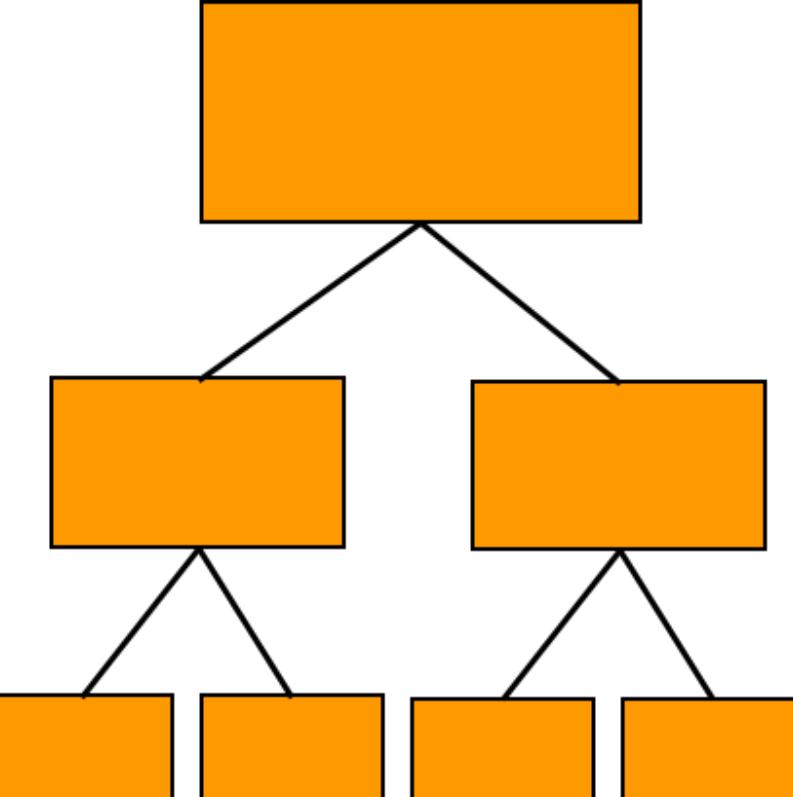


TABLE 2.1 Taxonomy of Common Multivariate Statistical Techniques			
Multiple predictors	Multiple outcomes	Multiple predictors and outcomes	
Exploratory	Confirmatory*	Exploratory	Confirmatory*
Degree of association			
Multiple regression	Hierarchical multiple regression	Factor analysis (monotonic)	Canonical correlation
Logistic regression	Hierarchical logistic regression	Hierarchical factor extraction	Confirmatory factor analysis (maximization of fit index)
Group differences <sup>a</sup>	ANOVA (planned comparisons)	Multidimensional scaling (specified dimensionality)	Conformatory factor analysis (minimization of specified dimensionality)
ANCOVA (post hoc comparisons)	ANCOVA (planned comparisons)	One-way MANOVA (post hoc comparisons)	Factorial MANOVA (post hoc comparisons)
Group membership	Hierarchical one-way discriminant analysis	One-way ANCOVA (post hoc comparisons)	Factorial MANCOVA (stepdowns and/or planned comparisons)
One-way discriminant analysis		One-way MANCOVA (post hoc comparisons)	Factorial MANCOV (stepdowns and/or planned comparisons)
			Hierarchical discriminant analysis

\* All listed techniques could conceivably be "confirmed" through cross-validation analysis using multiple samples. The confirmatory procedures listed above are the most common.

<sup>a</sup> There are no unique confirmatory applications for these inherently exploratory statistical techniques.

<sup>b</sup> ANOVA, analysis of variance; MANOVA, multiple analysis of variance; ANCOVA, analysis of covariance; MANCOVA, multiple analysis of covariance.

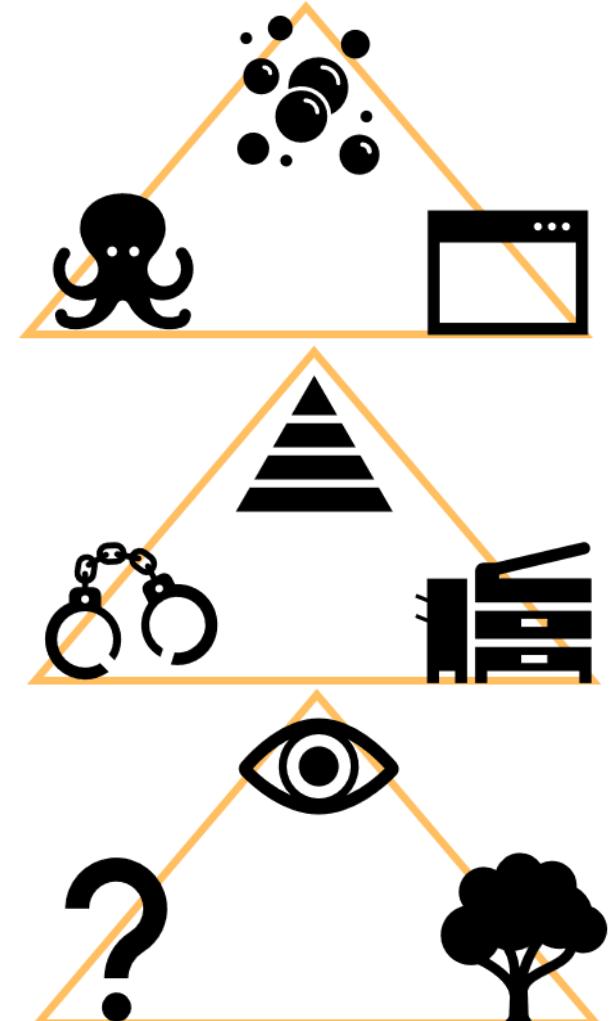


# Conclusions

- ⚒ Working with a biostatistician your best, but not only, option
- ⚒ If you struggle with a busy schedule, analyzing complex data, or getting your head around unknown data, there are a bundle of strategies to help you survive
- ⚒ Strategies are boilerplate code, flexible models, GUI usage, tiering methods, cheat sheets, multiple models, visualization first, data questionaries, and decision trees

**Please take the survey:**

Survey: [https://und.qualtrics.com/jfe/form/SV\\_9oWw3jNUcqDL8Gy](https://und.qualtrics.com/jfe/form/SV_9oWw3jNUcqDL8Gy)



# References

## Resources

- [1] <https://www.youtube.com/watch?v=nkkv96teOxs>
- [2] [https://med.und.edu/daccota/\\_files/docs/berdc\\_docs/mmgg\\_r\\_code.txt](https://med.und.edu/daccota/_files/docs/berdc_docs/mmgg_r_code.txt)
- [3] <https://www.youtube.com/watch?v=YtmloAwp2rl>
- [4] <https://www.rstudio.com/resources/cheatsheets/>
- [5] [https://med.und.edu/daccota/\\_files/docs/berdc\\_docs/statistical\\_software\\_toolkit\\_handout.docx](https://med.und.edu/daccota/_files/docs/berdc_docs/statistical_software_toolkit_handout.docx)
- [6] <http://www.ecostat.unical.it/Tarsitano/Didattica/LabStat2/Everitt.pdf>
- [7] <https://www.youtube.com/watch?v=eiivMNBZvw8>
- [8] [https://med.und.edu/daccota/\\_files/pdfs/berdc\\_resource\\_pdfs/stat\\_decision\\_tree.pdf](https://med.und.edu/daccota/_files/pdfs/berdc_resource_pdfs/stat_decision_tree.pdf)
- [9] <https://www.youtube.com/watch?v=kpmnFMznYt8>
- [10] <https://www.youtube.com/watch?v=YeKXdLFw0YI>

## Images

- [1] <https://d3gqasl9vmjfd8.cloudfront.net/83b73cac-f64d-4f7a-b6a5-78bbc6e6c559.png>
- [2] <https://plotly-book.cpsievert.me/images/plotlyGGally.gif>
- [4] <https://blogs.sas.com/content/graphicallyspeaking/files/2013/03/VBox4.png>
- [5] <https://i.pinimg.com/originals/47/1f/29/471f291ba7a7cd81c2118fe37be00b05.jpg>
- [6] <https://xkcd.com/1195/>



## Acknowledgements

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